

Original Article**Role of Adenosine Deaminase in Diagnosis of Extrapulmonary Tuberculosis: Study from a Tertiary Care Hospital of North India**

Rukhsana Taj, Unairah Naqash, Shazia Mushtaq

Abstract:

Background: Tuberculosis has been reported worldwide. Mycobacterium tuberculosis (MTB) primarily affect the lungs but can eventually spread to any other organ resulting in extra pulmonary tuberculosis (EPTB). The high morbidity associated with EPTB necessitates rapid development of sensitive tests. Adenosine Deaminase (ADA) is a proven biochemical marker with several advantages over conventional and molecular methods having high sensitivity and specificity for diagnosing TB.

Aims and Objective: To evaluate the sensitivity and specificity of Adenosine Deaminase (ADA) in diagnosis of extrapulmonary tuberculosis from different body fluids.

Methods: The current study was carried out on 190 specimens collected from different body fluids in order to rule out extrapulmonary tuberculosis. The smears were confirmed for presence of tubercular bacilli through ZN staining, LJ Media (Gold standard) and GeneXpert (CBNAAT). Simultaneously the ADA estimation was done from supernatant and readings were taken and expressed in IU/L.

Results: Out of the total 190 samples taken 31 (16.3%) samples tested positive for TB on culture media, 30 (15.8%) through GeneXpert and 2 (1.1%) on ZN staining. Through ROC analysis ADA estimation achieved the sensitivity of 100% in pleural, ascitic and synovial fluid. In CSF only 94.1% of sensitivity was achieved through ROC analysis. The specificity achieved was 100% for synovial fluid; however it reached to a maximum of 93.2% for pleural fluid, 95% for CSF and 96.4% for ascitic fluid.

Conclusion: Although conventional culture methods and GeneXpert are established diagnostic tools for identification of TB; the ADA levels estimation showed high sensitivity and specificity for diagnosis of tuberculosis in our study.

JK-Practitioner 2023;28(3-4):68-73**Introduction:**

Tuberculosis has been found to infect all age groups and its occurrence has been reported worldwide. The causative organism Mycobacterium tuberculosis (MTB) is the 13th leading cause of death world-wide.[1] The bacteria primarily affect the lungs but can eventually spread to any other organ directly or via lymphatics or hematogenous routes, resulting in extra pulmonary tuberculosis (EPTB).[2] In India 15%-20% of TB cases were reported to have TB involving extrapulmonary sites. 50% cases were seen in HIV positive individuals. [3] Extrapulmonary TB was seen mostly in adults above 44 years and children below 14 years of age.[4] Clinical presentation in EPTB is variable, depending on the organ involved, host immune response and the degree of tissue damage that has occurred. [5] Common disease manifestations include meningitis, lymphadenitis, pleuritis, pericarditis, peritonitis cutaneous, musculoskeletal, abdominal, genitourinary and miliary forms of tuberculosis.[6] The most frequent extrapulmonary sites of disease seen in India were extra thoracic lymph nodes (35%), pleural tuberculosis (20%), abdominal (14%) bone and joint tuberculosis (10%), genitourinary (5%), central nervous system (4.5%).[7]

The high morbidity associated with extra pulmonary tuberculosis (EPTB) necessitates rapid development of sensitive tests, the preliminary aim being to identify the presence of mycobacteria tuberculosis. Although a variety of tests including traditional,

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Keywords

ADA, Diagnostic Accuracy, Extrapulmonary Tuberculosis

Conventional culture-based techniques, Molecular Methods and Adenosine deaminase assay are available but the current study was limited to determine the diagnostic accuracy of ADA levels and a comparison to that of conventional and molecular methods for diagnosis of tuberculosis.

ADA is a proven biochemical marker that offers several advantages of being simple, rapid, low cost and easy to perform in most clinical laboratories.[8] The sensitivity and specificity of ADA in the different body fluids including those of pleural, pericardial, ascitic fluid etc has been ascertained in many studies.[9] In lieu of above facts the current study was undertaken to investigate the accuracy of ADA levels over other diagnostic approaches, wherein the specimens were collected from those of pleural fluids, ascitic fluids, synovial fluids, cerebrospinal and pericardial fluids for the diagnosis of EPTB. To evaluate the sensitivity and specificity of Adenosine Deaminase (ADA) in diagnosis of extrapulmonary tuberculosis from different body fluids.

Materials and Methods

This cross-sectional study was conducted at Postgraduate Department of Microbiology Govt. Medical College, Srinagar for a period of 18 months. A total of 190 specimens were collected from different body fluids like pleural fluid, ascitic fluid, cerebrospinal fluid (CSF), synovial fluid and pericardial fluid under all proper aseptic precautions from patients suspected of extra pulmonary tuberculosis. Specimens were collected in sterile, leak-proof, disposable, and appropriately labelled containers without fixatives and were transported to laboratory immediately. By keeping in consideration, the RNTCP guidelines for grading of ZN-stained smears, the differential staining technique (Ziehl - Neelson) was done for proper visualization and separation of acid-fast bacilli through microscope. Further, the specimens were concentrated by sedimentation in a refrigerated centrifuge at 3000 g for 30 minutes. The sediments were used for preparation of smears and were inoculated with 2-4 drops (0.2ml - 0.4ml) of centrifuged sediment on Lowenstein Jensen (LJ) media and were incubated at 35°C in the dark atmosphere and high humidity. Culture media was incubated in slanted position with screw caps and most isolates appeared between 3 and 6 weeks of incubation. After 8 weeks of incubation,

negative cultures (those showing no growth) were reported, and the culture bottles were discarded. The sediments were further analysed through GeneXpert /MTBRIF which is a rapid, nested real-time PCR for diagnosis of tuberculosis and drug resistance. It is a cartridge based nucleic acid amplification test (CBNAAT) which simultaneously detects DNA of Mycobacterium tuberculosis complex (MTBC) and resistance to rifampicin (RIF) in less than 2 hours. The supernatant was used for detecting adenosine deaminase (ADA) levels at the Department of Biochemistry through calorimetric method using the ADA assay kit in analyzer. Reading of ADA levels in different body fluids was done and ADA level was expressed in IU/L. The whole procedure was carried out in the biological safety cabinet.

Data Collection

The detailed history including chief complaints, co morbidities, history of contact with patients of tuberculosis and diagnostic results were entered on a self devised proforma. The data was entered in excel and analysed using SPSS version 20.0.

Ethical clearance

The ethical clearance for the study was granted by Ethical Clearance committee of Government Medical College, Srinagar.

Results:

The current study was conducted to rule out the efficiency of elevated Mean ADA levels in suspected cases along with a comparison to different multimodalities for the diagnosis of extrapulmonary tuberculosis. (Table:1) below shows the summarized report of different body fluids wherein a total of 190 samples were investigated for EPTB, out of which 64 (33.7%) samples were of pleural fluid, 57 (30.0%) of CSF, 33 (17.4%) of ascitic fluid, 32 (16.8%) of synovial fluid and 4 (2.1%) of pericardial fluid.

Outcome of culture on LJ media is shown in Table 1. In pleural fluid 64 samples were inoculated on LJ media and growth was seen in 5 (7.8%) samples and the same samples also tested positive on Genexpert. In 57 CSF samples growth was seen on 17 (29.8%) samples and same tested positive on GeneXpert. After inoculating 33 samples of ascitic fluid on LJ media 5 (15.2%) samples were positive and same tested also positive on GeneXpert. Four samples of synovial fluid tested positive on LJ

Table:1 Exudate types and their outcome on different diagnostic modalities.

Type of fluid	Total Samples	Culture Positive	GeneXpert Positive	ZN Staining Positive
Pleural Fluid	64	5 (7.8%)	5 (7.8%)	0 (0.0%)
CSF	57	17 (29.8%)	17 (29.8%)	1 (1.8%)
Ascitic Fluid	33	5 (15.2%)	5 (15.2%)	0 (0.0%)
Synovial Fluid	32	4 (12.5%)	3 (9.4%)	1 (3.1%)
Pericardial Fluid	4	0 (0.0%)	0 (0.0%)	0 (0.0%)
Total	190	31 (16.3%)	30 (15.8%)	2 (1.1%)

Type of sample & ADA Cutoff	No. of Samples	Overall Mean ADA	Std. Deviation	Mean \pm SD ADA levels among Culture Positives	Mean \pm SD ADA levels among Culture Negatives	P-value
Pleural Fluid	64	35.2077	46.57159	100.40 \pm 22.490	29.68 \pm 20.858	<0.001
CSF	57	18.6335	28.81398	51.00 \pm 34.808	4.877 \pm 1.660	<0.001
Ascitic Fluid	33	35.6430	56.71333	129.00 \pm 103.114	18.97 \pm 8.293	<0.001
Synovial Fluid	32	22.2941	20.77581	73.25 \pm 19.653	15.01 \pm 10.632	<0.001
Pericardial Fluid	4	17.0750	10.49170	-	17.07 \pm 10.491	-

media and 3 of them were positive on GeneXpert also. Only 1 sample each from CSF and synovial fluid tested positive for TB on ZN staining. All the 4 pericardial fluid samples were negative on all testing modalities.

Table 2 shows comparison of mean levels of ADA among tuberculosis positive and negative patients in different body fluids. In pleural fluid the Mean ADA level among culture positive samples was 100.4 \pm 22.490 and among culture negative samples was 29.68 \pm 20.858. The mean ADA activity among

culture positive CSF samples was 51.00 \pm 34.808 and among the culture negative samples it was 4.877 \pm 1.660. In ascitic fluid mean ADA level was 129.00 \pm 80.114 among culture positive subjects and among culture negative mean ADA was 18.97 \pm 8.293. In synovial fluid the mean ADA level among culture positive samples was 73.25 \pm 19.653 and in culture negative samples it was 15.01 \pm 10.632. The mean difference was statistically significant among positive and negative samples of pleural fluid, CSF, ascetic fluid and synovial fluid [p value (<0.001)].

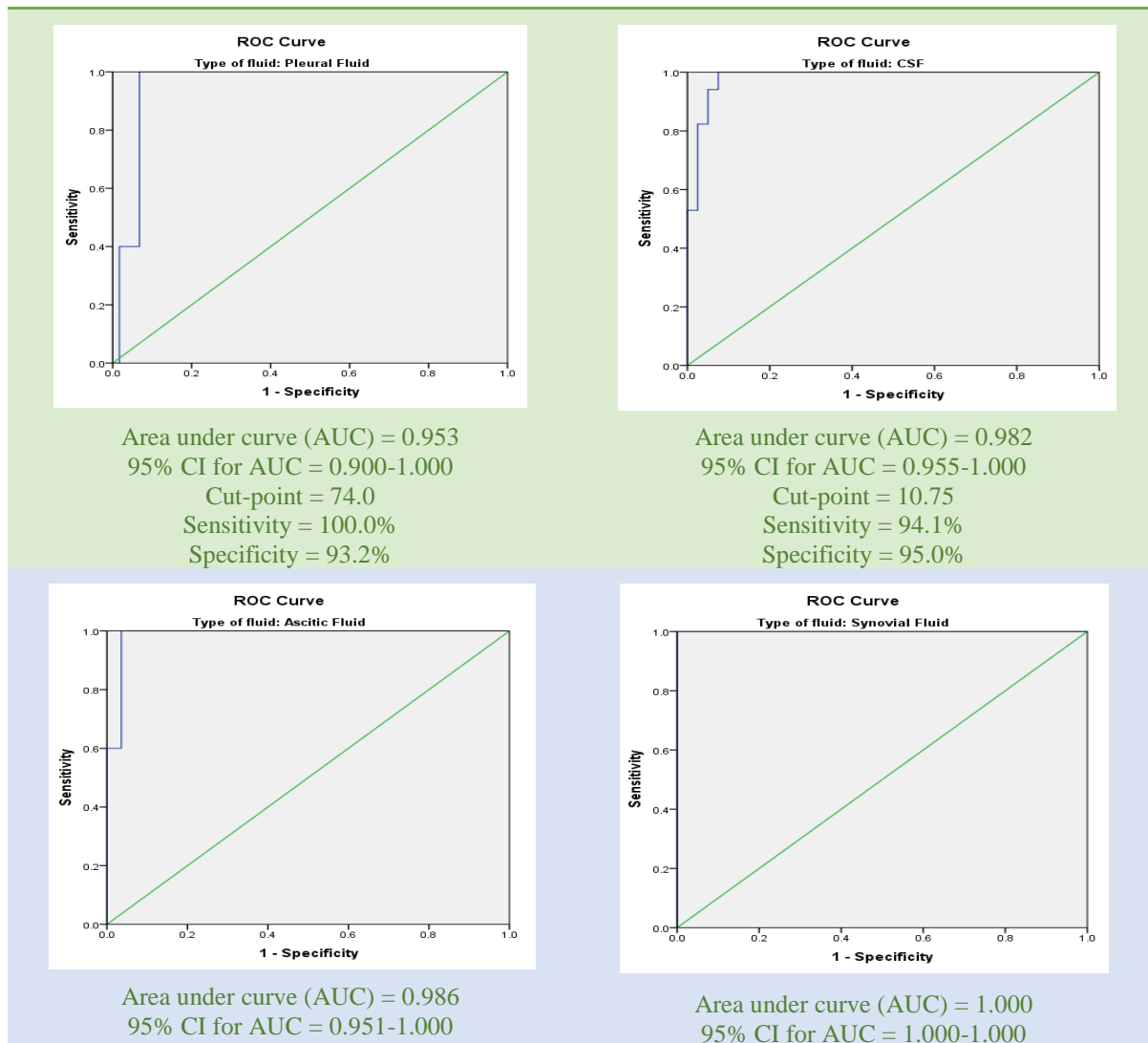
Type of fluid		Culture on LJ media		Sensitivity	Specificity	PPV	NPV	
		Positive	Negative					
Pleural Fluid	ADA Levels (Cut off 40U/L)	Positive	5	10	100.0%	81.4%	31.2%	100.0%
		Negative	0	49				
CSF	ADA Levels (Cut off 10U/L)	Positive	16	2	94.1%	95.0%	88.9%	97.4%
		Negative	1	38				
Ascitic Fluid	ADA Levels (Cut off 40U/L)	Positive	5	2	100.0%	92.9%	71.4%	100.0%
		Negative	0	26				
Synovial Fluid	ADA Levels (Cut off 40U/L)	Positive	4	0	100.0%	100.0%	100.0%	100.0%
		Negative	0	28				
Pericardial Fluid	ADA Levels (Cut off 40U/L)	Positive	0	0	-	100.0%	-	100.0%
		Negative	0	4				
Total	ADA Levels	Positive	48	17	94.1%	92.6%	73.8%	98.6%
		Negative	3	213				

Performance characteristics of ADA in different body fluids (at conventional cut off) when culture results were taken as gold standard is shown in **Table 3**. The sensitivity of ADA for the diagnosis of tuberculosis on pleural fluid was found to be 100% and specificity was 81.4%. In CSF the sensitivity and specificity of ADA levels in the diagnosis of tuberculosis was found 94.1% and 95.0% respectively. In case of ascitic fluid the sensitivity of ADA for the diagnosis of TB in ascitic fluid is 100% and specificity was 92.9%. Synovial fluid showed complete correspondence between ADA levels and culture positive growths, hence the sensitivity and specificity were calculated at 100% for both.

Table 4 shows performance characteristics of ADA in different body fluids with geneXpert as gold standard. The sensitivity of ADA for the diagnosis of tuberculosis in pleural fluid was found to be 100% and specificity was 81.4%. In CSF the sensitivity and specificity of ADA levels in the diagnosis of tuberculosis was found 94.1% and 95.0% respectively. In case of ascitic fluid the sensitivity of ADA for the diagnosis of TB in ascitic fluid is 100% and specificity was 92.9%. The ADA levels in synovial fluid showed sensitivity and specificity 100% & 96.6%, respectively.

Table 4: Performance characteristics of ADA Levels in different types of fluids as compared to GeneXpert in our study population								
Type of fluid			GeneXpert		Sensitivity	Specificity	PPV	NPV
			Positive	Negative				
Pleural Fluid	ADA Levels (Cut off 40U/L)	Positive	5	10	100.0%	81.4%	31.2%	100.0%
		Negative	0	49				
CSF	ADA Levels (Cut off 10U/L)	Positive	16	2	94.1%	95.0%	88.9%	97.4%
		Negative	1	38				
Ascitic Fluid	ADA Levels (Cut off 40U/L)	Positive	5	2	100.0%	92.9%	71.4%	100.0%
		Negative	0	26				
Synovial Fluid	ADA Levels (Cut off 40U/L)	Positive	3	1	100.0%	96.6%	75.0%	100.0%
		Negative	0	28				
Pericardial Fluid	ADA Levels (Cut off 40U/L)	Positive	0	0	-	100.0%	-	100.0%
		Negative	0	4				

Fig. 1: ROC analysis of ADA in different body fluids when compared to culture (gold standard) for EPTB diagnosis.



Cut-point = 53.5
Sensitivity = 100%
Specificity = 96.4%

Cut-point = 45.5
Sensitivity = 100.0%
Specificity = 100.0%

Figure 1 shows receiver operating characteristic (ROC) of ADA in different body fluids. The area under curve achieved was 0.953 (0.900-1000) for ADA in pleural fluid. At a cutoff of 74.0 IU/L for ADA in pleural fluid the sensitivity and specificity achieved was 100% & 93.2%, respectively. In CSF the area under curve for ADA achieved was 0.982 (0.955-1000). At a cutoff of 10.75 IU/L for ADA in CSF the sensitivity and specificity achieved was 94.1% & 95.0%, respectively. In ROC analysis (ADA in ascitic fluid) the area under curve achieved was 0.986 (0.951-1000). At a cutoff of 53.5 IU/L for ADA in ascitic fluid the sensitivity and specificity achieved was 100% & 96.4%, respectively. **Figure 1** also shows receiver operating characteristic (ROC) of ADA in synovial fluid. The area under curve achieved was 1.000 (1.000-1000). At a cutoff of 45.5 IU/L for ADA in synovial fluid the sensitivity and specificity achieved was 100%.

Discussion:

The present study was conducted on 5 different types of body fluids including those of Pleural fluid, CSF, Ascitic fluid, synovial fluid and pericardial fluid in order to find out the utility of ADA levels in diagnostic accuracy of EPTB. Out of the total 64 samples taken, 5 samples were positive in both ADA and culture media while 49 were being found negative. The Sensitivity of ADA for the diagnosis of extrapulmonary tuberculosis was found to be 100% while the specificity attained was 81.4% on both culture and GeneXpert. This analysis showed a close resemblance with the studies of **San José et al. [10]** wherein they analysed the use of serum, pleural ADA and lysozyme in tuberculous pleurisy in Spain, and found that pleural ADA had a sensitivity of 100% and a specificity of 93% for the diagnosis of TB. **Muranishi et al. [11]** while working on measurement of ADA activity and tuberculostearic acid (TSA) in pleural effusions attained a sensitivity of 56% and specificity of 76% for the diagnosis of tuberculous pleuritis. However, the observed results were relatively low when compared with our data. Our results further showed a concordance with the studies of **De Oliveira et al. [12]** who concluded that the use of these tests in combination was a highly efficient diagnostic strategy of low cost that merits wider use. On ROC analysis of pleural ADA, at a cut off of 74.0 IU/L, 100% sensitivity and 93.2% specificity were achieved. The predictive value (cut-off) as reported by **Villena et al. [13]** in their study was 33 IU/L for ADA whereby a sensitivity of 90% and specificity of 85% was achieved for tuberculous effusion, however, this value was much lower than our study.

The mean ADA activity among culture positive samples in CSF was 51.00 ± 34.808 and among culture negative samples was 4.877 ± 1.660 with a significant p value (<0.001). **Gambhir et al. [14]** reported mean ADA of 9.6 ± 4.1 U/L in 36 patients with tubercular meningitis, which was lower than the mean ADA in our study (18.6335 ± 28.813) however, the fact may be attributed to the inclusion of more adults in our study. Upon both culture media and GeneXpert, the sensitivity and specificity of ADA levels in the diagnosis of EPTB came out to be 94.1% and 95.0% respectively. The results went in a complete harmony with the studies of **Choi et al. [15]** who observed ADA activity in CSF of 36 TBM patients and reported that at a cut-off of 7 IU/L, the sensitivity of the test for TBM group as compared to aseptic meningitis group was 83% and the specificity was 95%. Same sensitivity and specificity like that on culture media and GeneXpert was achieved upon ROC analysis, at a cut-off of 10.75 IU/L for ADA in CSF. However, **Corral et al. [16]** used ROC curve analysis and suggested a cut off value of 8.5 U/L for the diagnosis of TBM with 57% sensitivity and 87% specificity. For ascitic fluid the mean ADA level observed among culture positive subjects was 129.00 ± 80.114 and among culture negative subjects was 18.97 ± 8.293 . The sensitivity of ADA for the diagnosis of EPTB in ascitic fluid when compared with LJ media (Gold Standard) and GeneXpert was 100% and the specificity was 92.9%. Upon ROC analysis area under the curve achieved was 0.986 (0.951-1000) and at a cut-off of 53.5 IU/L for ADA in ascitic fluid the sensitivity and specificity achieved was 100% & 96.4%, respectively for diagnosis of peritoneal tuberculosis. However, **Hillenbrand DJ et al. [17]** obtained a sensitivity of only 30% in their study and concluded that the ADA is inferior in cirrhosis as a diagnostic method for peritoneal TB. Furthermore, they inferred that this could be because of a positive correlation between ADA and ascitic fluid total protein as cirrhotic patients generally have low ascitic fluid total protein. In a study conducted by **Dahale AS et al. [18]** the ADA had very high sensitivity (93%) and specificity (94%) at a cut-off of 41.1 IU/L. They also showed the presence of elevated ADA levels in the cirrhotic peritoneal TB group than that of cirrhotic non-peritoneal group and all other types of ascites patients in the control group. Similarly, 4 samples of synovial fluid out of 32 samples taken were found positive on Culture media and there was a complete correspondence between ADA levels and culture growth, hence the sensitivity and specificity calculated was 100% for both. Upon ROC analysis the area under curve achieved was 1.000

(1.000-1000) and at a cut-off of 45.5 IU/L, the sensitivity and specificity achieved was 100%. **Gupta VK et al. [19]** while studying Musculo-skeletal disease observed a positive predictive value of 85.71% and a negative predictive value of 66.67%, however, the predictive values being lower than that found in our study.

Conclusion:

Although culture and GeneXpert are good diagnostic tools for identification of TB yet estimation of ADA levels in patients suspected of TB especially EPTB is inexpensive, rapid and efficient with high sensitivity and specificity. It is also efficient in differentiating tubercular pulmonary infections from non-tubercular pulmonary infections thereby, suggesting that the test should be included in routine investigations in patients suspected of tuberculosis.

References:

1. Global tuberculosis report 2021. Geneva: World Health Organization, 2021. Available from <https://www.who.int/publications/i/item/9789240037021>
2. Sharma SK, Mohan A. Extra pulmonary tuberculosis. *Indian J Med Res.* 2004 Oct;120(4):316-53. PMID: 15520485
3. Revised National TB control programme. Technical and operational guidelines for Tuberculosis control in India. 2020.
4. Rai DK, Pandey S. A hospital based cross sectional study on clinico-demographic characteristic of extra pulmonary tuberculosis cases coming to a tertiary hospital of Bihar. *Indian J Community Med.* 2018;43:122-3.
5. Mbuh TP, Ane-Anyangwe I, Adeline W, Pokam BDT, Meriki HD, Mbacham WF. Bacteriologically confirmed extra pulmonary tuberculosis and treatment outcome of patients consulted and treated under program conditions in the littoral region of Cameroon. *BMC Pulm Med.* 2019;19(17). <https://doi.org/10.1186/s12890-018-0770-x>
6. Lin JN, Lai CH, Chen YH, Risk factors for extra-pulmonary tuberculosis compared to pulmonary tuberculosis. *Int. J Tuberc Lung Dis.* 2009;13:620-625.
7. Kumar SA, Brahmachari S, Pathak P, Kumar R, Sainia T, Patel U, Mandil A. Clinico-Epidemiological Profile of Extra-pulmonary Tuberculosis in Central India. *Int J Med Res Rev.* 2015 Mar ;3(2):223-30.
8. Riantawan P, Chaowalit P, Wongsangiem M, Rojanarawee Wong P. Diagnostic value of pleural fluid adenosine deaminase in tuberculous pleuritic with reference to HIV coinfection and a Bayesian analysis. *Chest.* 1999; 116:97-103.
9. Barau R and Hossain M. Adenosine deaminase in diagnosis of tuberculosis: a review. *Anwer Khan Mod. Med Coll J.* 2014;5:43-8.
10. Jose MES, Valdes L, Saavedra MJ, De-Vega JM, Alvarez D, Viñuela J, et al. Lymphocyte population in tuberculosis pleural effusion. *Ann Clin Biochem.* 1999 Jul;36 (Pt 4):492-500. doi: 10.1177/000456329903600413.
11. Muranishi H, Nakashima M, Hirano H, Saitoh T, Takahashi H, Tanaka K, Miyazaki M, et al. Simultaneous measurements of adenosine deaminase activity and tuberculo-stearic acid in pleural effusions for the diagnosis of tuberculous pleuritis. *Intern Med.* 1992;31(6):752-755. 20.
12. De Oliveira HG, Rossatto ER, Prolla JC. Pleural fluid adenosine deaminase and lymphocyte proportion: clinical usefulness in the diagnosis of tuberculosis. *Cytopathology.* 1994 Feb;5(1):27-32.
13. Villena V, Navarro-González JA, García-Benayas C, Manzanos JA, Echave J, López-Encuentra A, Arenas Barbero J. Rapid automated determination of adenosine deaminase and lysozyme for differentiating tuberculous and nontuberculous pleural effusions. *Clin Chem.* 1996 Feb;42(2):218-21.
14. Gambhir IS, Mehta M, Singh DS, Khanna HD. Evaluation of CSF-adenosine deaminase activity in tubercular meningitis. *J Assoc Physicians India.* 1999 Feb;47(2):192-4.
15. Choi SH, Kim YS, Bae IG, Chung JW, Lee MS, Kang JM, Ryu J, Woo JH. The possible role of cerebrospinal fluid adenosine deaminase activity in the diagnosis of tuberculous meningitis in adults. *Clin Neurol Neurosurg.* 2002 Jan;104(1):10-5.
16. Corral I, Queda C, Navas E, Martín-Dávila P, Pérez-Elías MJ, Casado JL, et al. Adenosine deaminase activity in cerebrospinal fluid of HIV-infected patients: limited value for diagnosis of tuberculous meningitis. *Eur J Clin Microbiol Infect Dis.* 2004 Jun;23(6):471-6. doi: 10.1007/s10096-004-1110-z. Epub 2004 May 13.
17. Hillebrand DJ, Runyon BA, Yasmineh WG, Rynders GP. The American Association for the Study of Liver Diseases. December 1996; 24(6):1408-1412.
18. Dahale AS, Puri AS, Sachdeva S, Agarwal AK, Kumar A, Dalal A, Saxena P. Reappraisal of the Role of Ascitic Fluid Adenosine Deaminase for the Diagnosis of Peritoneal Tuberculosis in Cirrhosis. *Korean J Gastroenterol* 2021;78(3):168-176.
19. Gupta VK, Mukherjee S, Dutta SK and Mukherjee P. Diagnostic evaluation of ascitic adenosine deaminase tubercular peritonitis. *J. Assoc. Physicians.* 1992;40: 387-389.